

Development of a Respiratory Pathogen Panel with an Automated High Throughput System - BioCode MDx-3000

Applied BioCode

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Abstract

Introduction

Acute respiratory tract infections (ARIs) are the most common causes of childhood morbidity and mortality worldwide, which accounts for about 30% of all childhood deaths in the developing world. However in developed countries, ARIs cause enormous direct and indirect health costs¹. Rapid identification of respiratory viruses can help in avoiding indiscriminate use of antibiotics as well as large healthcare costs. Using our proprietary Barcoded Magnetic Bead (BMB) technology, Applied BioCode is developing a multiplex molecular diagnostic assay for detection of respiratory pathogens in a high-throughput 96-well format. The BioCode Respiratory Panel is designed for detection of Influenza viruses (Influenza A: subtype H1N1 2009 Pandemic, H1 seasonal, H3, Influenza B), Parainfluenza viruses (type 1, 2, 3, 4A, 4B), RSV, human Metapneumovirus, Rhinovirus/ Enterovirus, Coronavirus (OC43, NL63, 229E, HKU1), Adenovirus and bacteria (*Bordetella pertussis*, *B. parapertussis*, *Mycoplasma pneumoniae*, *Chlamydophila pneumoniae*, and *Legionella pneumophila*).

1 Hinman, A. R. 1998. Global progress in infectious diseases control. Vaccine 16:1116-1121

Materials & Methods

Biocode MDx-3000 platform, an automated PCR and post-PCR sample handling and detection system in 96-well format was used for the study. Off board automated nucleic acid extraction system was used and pathogenic targets were amplified by one-step RT-PCR. PCR products were captured by target-specific probes coupled to barcoded magnetic beads (BMBs) and fluorescent signal was generated by incubation with a conjugate (Figure 1). Qualitative results were determined by Median Fluorescent Index (MFI) of fluorescent signals from analyte-specific BMBs (Figure 2).

Conclusions

The BioCode Respiratory Pathogen Panel run on BioCode MDx-3000 system, specifically and reproducibly detects viruses and bacteria known to cause upper respiratory infections. Combined, the automated system and molecular panel allows users to perform highly multiplexed molecular detection in a high-throughput, automated format with a simple workflow and minimal hands-on time.

- ❖ No cross reactivity was observed with organisms tested (Table 1).
- ❖ Preliminary LoD of the BioCode Respiratory Pathogen Panel is shown in Table 2.
- ❖ Inclusivity was tested with several relevant pathogens which were detected with high titer of pathogen (Table 3).
- ❖ Method comparison of 208 NPS showed 88% overall (and 92% without rhinovirus) positive agreement (Table 4 & 5).

The BioCode Respiratory Pathogen Panel is currently in development.
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Multiplex Respiratory Pathogen Panel

Viruses

- Influenza A
 - H1N1 2009pdm
 - H1 (seasonal)
 - H3
- Influenza B
- RSV A/B
- Human metapneumovirus
- Adenovirus
- Rhinovirus/ Enterovirus
- Parainfluenza virus type 1
- Parainfluenza virus type 2
- Parainfluenza virus type 3
- Parainfluenza virus type 4
- Coronavirus 229E
- Coronavirus HKU1
- Coronavirus NL63
- Coronavirus OC43

Bacteria

- *Bordetella pertussis*
- *Chlamydophila pneumoniae*
- *Mycoplasma pneumoniae*
- *Bordetella parapertussis*
- *Legionella pneumophila*

Barcoded Magnetic Bead (BMB) Technology

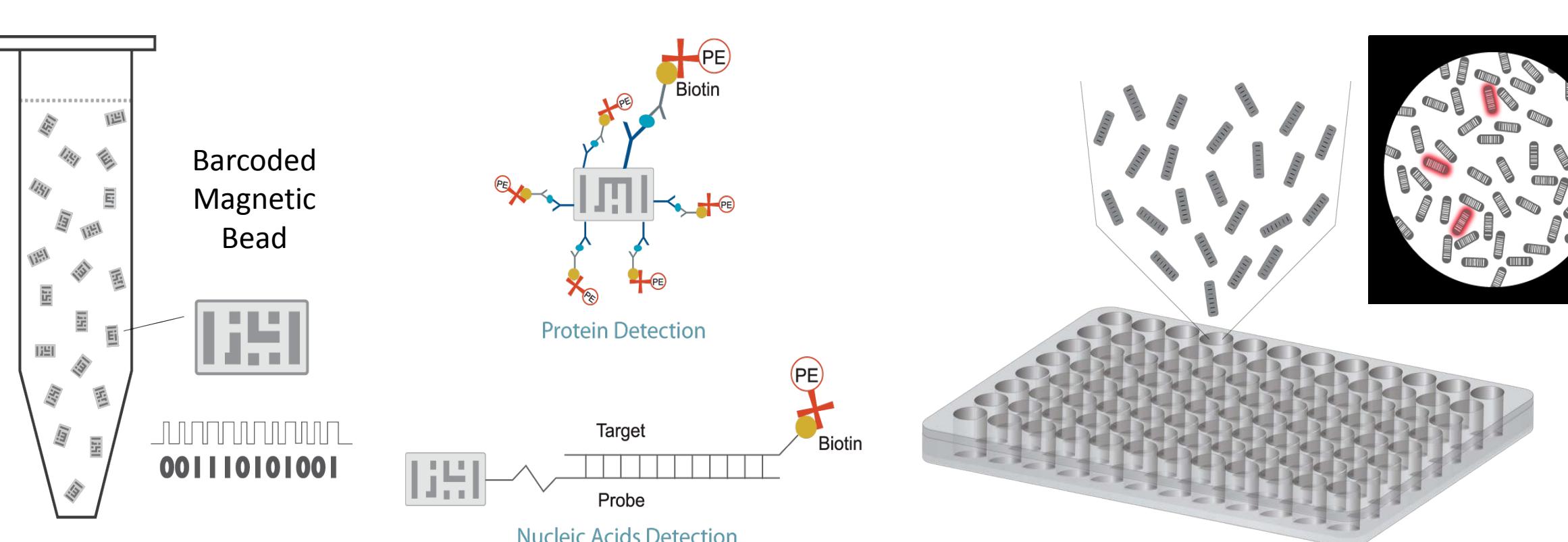


Figure 1. Barcoded Magnetic Beads (BMBs) are coupled to proteins or nucleic acids probes and used for target capture in microtiter plates. In the BioCode Respiratory Pathogen Panel, biotinylated PCR product is captured by target-specific nucleic acid probes coupled to BMBs then labeled by SA-PE for detection.

BioCode MDx-3000

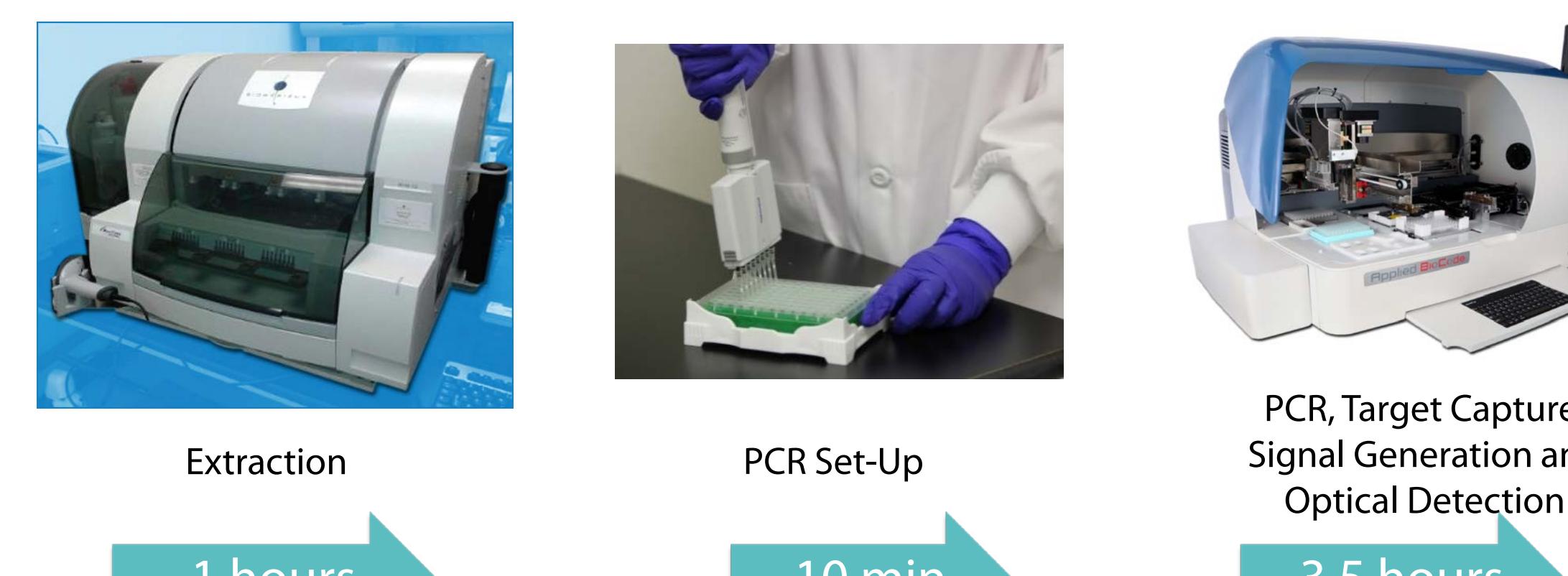
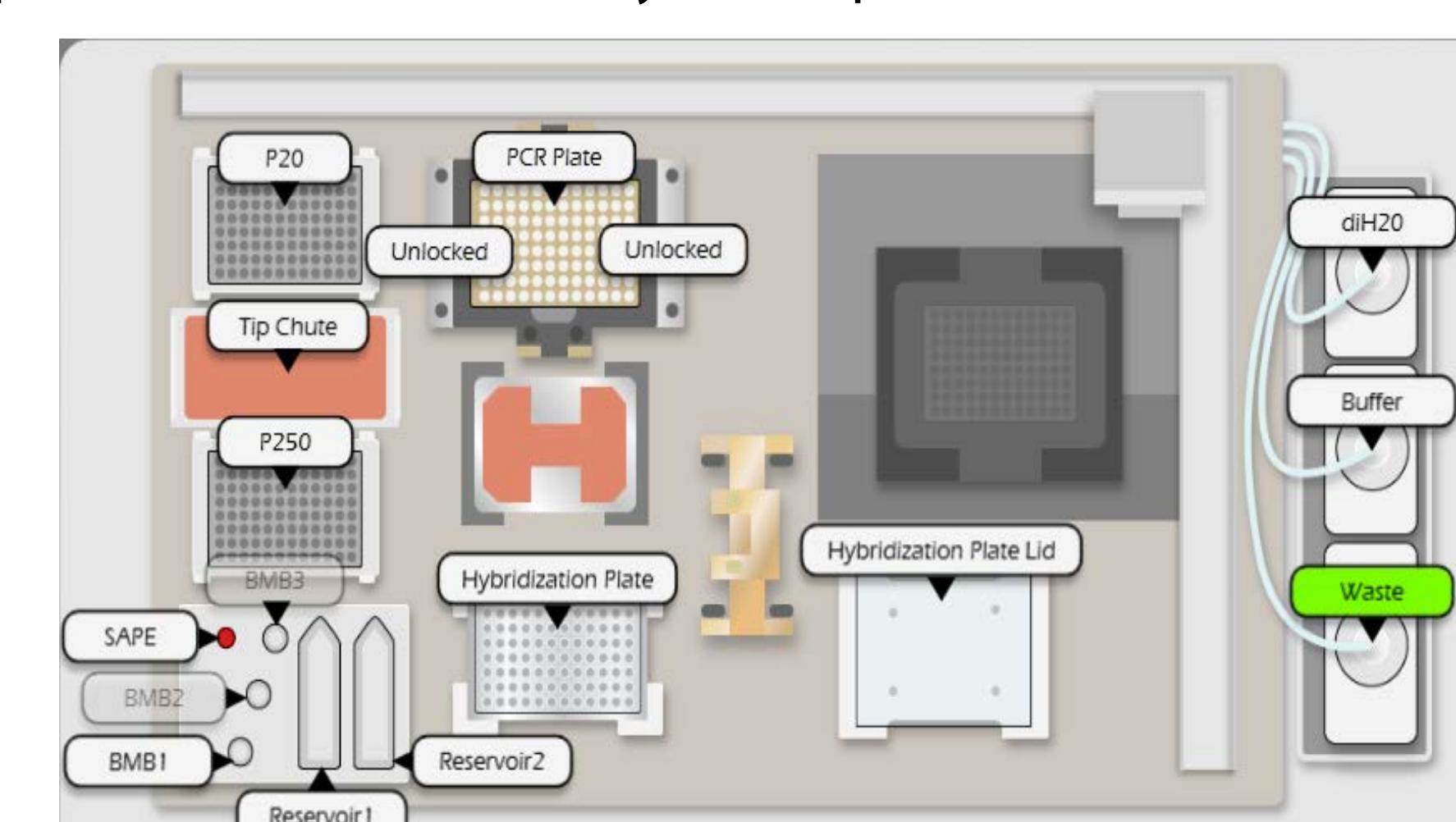


Figure 2. Workflow for BioCode Respiratory Pathogen Panel (top) and schematic for BioCode MDx-3000 deck layout (bottom). 192 samples in an 8 hour shift with minimal hands on time. Three different BioCode panels can be performed simultaneously in one plate.



Cross Reactivity Studies

Table 1. Microorganisms tested for cross reactivity

Bacteria and Fungi		
<i>E. Coli</i> 35328	<i>Pseudomonas aeruginosa</i>	<i>Candida albicans</i>
<i>E.coli</i> BAA-1431	<i>Staphylococcus aureus</i> 3300	<i>Klebsiella pneumoniae</i>
<i>Haemophilus influenzae</i>	<i>Staphylococcus epidermidis</i>	<i>Moraxella catarrhalis</i>
<i>Lactobacillus acidophilus</i>	<i>Streptococcus intermedius</i>	<i>Neisseria elongata</i>
<i>Lactobacillus reuteri</i> Strain	<i>Streptococcus pyogenes</i>	<i>Chlamydia trachomatis</i>
<i>Neisseria gonorrhoeae</i>	<i>Streptococcus salivarius</i>	<i>Lactococcus lactic</i>
<i>Acinetobacter baumannii</i>		

Viruses		
Epstein-Barr Virus (EBV)	Bocavirus	Cytomegalovirus (CMV)
Herpes Virus	Mumps	

➤ No cross reactivity was observed with bacteria ($\geq 10^6$ CFU/mL), viruses ($\geq 10^5$ TCID₅₀/mL)

Preliminary Limit of Detection (LoD)

Table 2. Limit of Detection (LoD) for the BioCode Respiratory Pathogen Panel performed on the BioCode MDx-3000 system

Organism	Source	Preliminary LoD
Viruses		
Human Parainfluenza 1	BEI NR-48680	≤ 1.25 TCID ₅₀ /mL
Human Parainfluenza 2	BEI NR-3229	≤ 10 TCID ₅₀ /mL
Human Parainfluenza 3	Zeptometrix 0810016CF	≤ 1.25 TCID ₅₀ /mL
Human Parainfluenza 4	Zeptometrix 0810060CF	≤ 1.25 TCID ₅₀ /mL
Rhinovirus	Zeptometrix 0810012CFN	≤ 10 TCID ₅₀ /mL
RSV A & B	Zeptometrix 0810040ACF	≤ 1.25 TCID ₅₀ /mL
Coronavirus OC43	Zeptometrix 0810024CF	≤ 1.25 TCID ₅₀ /mL
Coronavirus 229E	Zeptometrix 0810229CF	≤ 1.25 TCID ₅₀ /mL
Coronavirus NL63	Zeptometrix 0810228CF	≤ 1.25 TCID ₅₀ /mL
Coronavirus HKU1	IDT-Ultramer	≤ 100 copies/mL
Human Metapneumovirus	Zeptometrix 0810161CF	≤ 100 TCID ₅₀ /mL
Influenza A H3	Zeptometrix 0810252CF	≤ 10 TCID ₅₀ /mL
Influenza A H1 subtype	Zeptometrix 0810036CF	≤ 1.25 TCID ₅₀ /mL
Influenza A H3 subtype	Zeptometrix 0810252CF	≤ 10 TCID ₅₀ /mL
Influenza A H1N1 2009pdm	IDT-Ultramer	≤ 100 copies/mL
Influenza B	Virapur C1320H	≤ 10 TCID ₅₀ /mL
Adenovirus	Zeptometrix 0810050CF	≤ 1 TCID ₅₀ /mL
Bacteria		
<i>Bordetella pertussis</i>	Zeptometrix 0801460	≤ 1.25 CFU/mL
<i>Bordetella parapertussis</i>	Zeptometrix 0801461	≤ 2.5 CFU/mL
<i>Chlamydophila pneumoniae</i>	ATCC VR-1360D	≤ 100 copies/mL
<i>Legionella pneumoniae</i>	ATCC 33152D-5	≤ 100 copies/mL
<i>Mycoplasma pneumoniae</i>	Zeptometrix 0801579	≤ 1.25 CFU/mL

Method Comparison Study

Table 5. Clinical Agreement: BioCode RP Panel vs Comparator

Overall Agreement	GenMark RVP		
	Pos	Neg	Total
BioCode RP Panel	165	0	165
	23	20	43
	188	20	208
		Positive Agreement	88%
		Negative Agreement	100%

Inclusivity Study

Table 3. Organisms detected for Inclusivity of the BioCode Respiratory Pathogen Panel

Organisms Detected
Influenza A/Denver/1/1957
Influenza A/Fort Monmouth/01/1947
Influenza A/New Jersey/8/1976
Influenza A/Taiwan/42/06
Influenza A/Singapore/63/04
Influenza A/ Aichi/2/68
Influenza A/Port Chalmers/1/1973
Influenza A/Switzerland/9715293/2013
Influenza A/Wisconsin/15/2009 H3N2
Influenza A/Beijing/262/95
Influenza A/Brisbane/59/2007 H3N2
Influenza A/Uruguay/706/07
Influenza A/New Caledonia/20/99
Influenza A/Solomon Islands/03/2006
Influenza A/H3N2/Hong Kong
Influenza A/H3N2/Victoria/3/1975
Influenza B/Maryland/1/59
Influenza B/PHUKET/3073/2013
Influenza B/GL/1739/55
Influenza B/Hong Kong/5/1972
Influenza B/Brisbane/60/2008
Influenza B/Florida/04/06
Influenza B/Christ Church
Influenza B/Sydney/507/2006
Influenza B/OHIO/01/2005
Influenza B/Malaysia/2506/04
Influenza B/Texas/06/2011
Influenza B/Nevada/03/2011
<i>Bordetella pertussis</i> A747
<i>Bordetella pertussis</i> E431
<i>Bordetella pertussis</i> A639

Method Comparison Study

Table 4. Detection of target pathogens: BioCode Respiratory Panel vs previously reported test results

Target Pathogens	Positive Results reported by	
	BioCode RP Panel	GenMark RVP
Influenza A (not subtyped)	20	20
Influenza A H3	11	11
Influenza A H1N1 2009pdm	14	18
Influenza A H1	2	2
Influenza B	28	31
RSV	38	42
Rhinovirus/ Enterovirus	17	27
Metapneumovirus	20	